

Westmead Research Hub Imaging Resources updated 10/4/08

Items are listed with:

Description + (Name on CHW Booking System) or (Name on WMI Booking System)

Children's Hospital Westmead

The link to the CHW booking system is:

<http://www.chw.edu.au/research/hub/mrbs/day.php>

Leica Confocal Microscope (Confocal)

Location: Room 444873 Level 4 Research Building (CHW)

Contact: Laurence Cantrill (53087; laurenc@chw.edu.au)

Description: Leica TCS SP2 confocal microscope with an upright and inverted microscope (which also has live cell imaging capability, with heated CO₂ chamber). Blue argon, and green and red helium-neon lasers. Information available online at

http://www.chw.edu.au/research/hub/microscopy/equip_info.htm#confocal

Digital Camera and Imaging Software (Live Cell Imaging Camera)

Location: Room 444873 Level 4 Research Building (CHW)

Contact: Laurence Cantrill (53087; laurenc@chw.edu.au) Description: SPOT Camera attached to Leica DMIRBE inverted fluorescent microscope. Shuttered fluorescent and bright field sources allow time lapse imaging of either illumination. Image Pro 5 imaging software.

http://www.chw.edu.au/research/hub/microscopy/equip_info.htm#confocal

Fluorescent Microscopes (x2) (SPOT microscope & DC500)

Location: Room 434876 Level 3 Research Building (CHW)

Contact: Laurence Cantrill (53087; laurenc@chw.edu.au) Description: Olympus BX50 with Jenoptik digital camera and ProgRes V2.5 image capture software & Leica DMLA with QiCam camera and Q Capture image capture software. Information available online at:

http://www.chw.edu.au/research/hub/microscopy/equip_info.htm#optical

Dissecting Microscope (Dissecting Microscope)

Location: Room 434876 Level 3 Research Building (CHW)

Contact: Laurence Cantrill (53087; laurenc@chw.edu.au)

Description: Leica MZ8 Dissecting Microscope with Leica Schott KL1500 cold light source, DC500 camera and Image Manager (IM50) image capture software.

Inverted Fluorescent Microscope/Digital Camera

Location: Level 3 South Wing (CHW)

Contact: Laurence Cantrill (53087; laurenc@chw.edu.au)

Description: DP12 Olympus digital camera attached to inverted light microscope (with phase and fluorescence- blue and green excitation filters).

Inverted Light Microscope/ Digital Camera

Location: TC Room 1 Level 3 Research Building (CHW)

Contact: Laurence Cantrill (53087; laurenc@chw.edu.au)

Description: DP12 Olympus digital camera attached to inverted light microscope (with phase).

PALM Laser Capture Microdissection system (LCM)

Location: Room 434871 Level 3 Research Building (CHW)

Contact: Laurence Cantrill (53087; laurenc@chw.edu.au)

Description: PALM Robot-MicroBeam Research system - broad range of applications including single cell DNA and mRNA analysis and separation of single cells from cultures. Information available online at:

http://www.chw.edu.au/research/hub/microscopy/equip_info.htm#laser

ORU Live Cell Imaging Facility

Location: Room 444874 Level 4 Research Building (CHW)

Contact: Geraldine O'Neill (53116; geraldio@chw.edu.au)

Description: Olympus IX81 motorised inverted fluorescent microscope with motorised stage and Metamorph image collection and analysis software. Hamamatsu Photonics Cooled CCD camera. Blue/Green/Red/Far Red filter sets, fully enclosed chamber, heating and CO₂

Aperio CS Virtual Microscopy Suite

Location: Room 444874 Level 4 Research Building (CHW)

Contact: Daniel Catchpoole (51205; DanielC@chw.edu.au)

Description: Automated slide imager with capture and storage software

Beta counter and x-ray processor

Location: Level 3 corridor Research Building (CHW)

Contact: Matt Laver (53091; matthel1@chw.edu.au)

Description: Radiation assays + auto radiography

Westmead Hospital

Scanning Electron Microscope

Location: Institute of Dental Research Level 2 (Westmead Dental School)

Contact: Mara Cvejic (57412)

Description: Joel 840

Transmission Electron Microscope

Location: Level 1 Electron Microscope Lab (ICPMR)

Contact: Ross Boadle (Senior Scientist) (57783 Page: 01266)

Description: Phillips CM10 TEM. The laboratory is well equipped for conventional (resin) and low temperature techniques, the latter including cryofixation, and cryosectioning and freeze substitution. Both morphological and immunolabelling techniques are routinely used.

Scanning Transmission Electron Microscope

Location: Level 1 Electron Microscope Laboratory (ICPMR)

Contact: Ross Boadle (Senior Scientist) (57783 Page: 01266)

Description: Phillips CM120 BioTWIN (S) TEM with x-ray microanalysis.

Confocal Microscope

Location: Room 4.2288 Level 4. Centre for Oral Health (Westmead Hospital)

Contact: Professor Neil Hunter (58762), Derrick Dehati (58772), Sheryl Chapel (58770)

Description: Optiscan Confocal with an Argon Laser.

Video/VCR Microscope

Location: Anatomical Pathology (Westmead Hospital)

Contact: Bill Sinai (57774)

Description: Video attached to microscope, record to VCR. No PC capture card.

Image Analyser

Location: Lab 2155 Clinical Sciences Area (Westmead Hospital)

Contact: Wayne Hawthorne (57365; wayneh@med.usyd.edu.au)

Description: Optimas system (version 6.5.1) Computer image analysis system, uses a light box to image directly to a digital camera

Olympus BX60 Fluorescence Microscope and Spot camera

Location: Lab 2154 Clinical Sciences Area (Westmead Hospital)

Contact: Hong Yu (59515; hong_yu@wmi.usyd.edu.au)

Description: Olympus BX60 with newly upgraded phase contrast condenser; DAPI/FITC/TexasRed filter sets, awaiting computer replacement

Westmead Millennium Institute

The link to the WMI booking system is:

<http://www.wmi-htc.org.au>

Olympus FV 1000 confocal laser scanning microscope (Olympus Confocal Microscope)

Location: Room 3048 Level 3 WMI Building

Contact: Hong Yu (59515; hong_yu@wmi.usyd.edu.au)

Description: Olympus IX-81 ZDC microscope with motorised stage, live cell imaging heated chamber and CO₂ Lasers: 405nm,473nm,559nm,633nm. Olympus FV10-ASW 1.7a capture software.

Imaging analysis PC (Image Analysis Workstation)

Location: Room 3048 Level 3 WMI Building

Contact: Hong Yu (59515; hong_yu@wmi.usyd.edu.au)

Description: Olympus FV10-ASW 1.7a analysis software.

Live Cell Imaging Microscope (Live Cell Imaging Microsc)

Location: Room 2019 Level 2 WMI Building

Contact: Hong Yu (59515; hong_yu@wmi.usyd.edu.au)

Description: Zeiss AxioVert 200M motorised microscope with motorised stage, fully-enclosed incubator with CO₂ supply and temperature controller, Zeiss AxioCam HRM digital camera, Zeiss AxioVision capture software. Blue/Green/Red/Far Red filter sets.

SPOT Microscope System

Location: Room 2019 Level 2 (WMI)

Contact: Hong Yu (59515; hong_yu@wmi.usyd.edu.au) Description: Fluorescent Olympus microscope with Spot digital camera and Spot slider software. UV, FITC, Texas Red and double excitation fluorescence filters for FITC and TexasRed

SPOT Microscope System (Leica SPOT 4)

Location: Room 3014 Level 3 (WMI)

Contact: Hong Yu (59515; hong_yu@wmi.usyd.edu.au) Description: Upright fluorescent Leica microscope with Spot digital camera and Spot software. UV, FITC, Texas Red filters.

Inverted Leica fluorescence microscope (Inverted Leica fluorescence microscope)

Location: Room 3014 Level 3 (WMI)

Contact: Hong Yu (59515; hong_yu@wmi.usyd.edu.au) Description: inverted fluorescent Leica microscope with Leica digital camera and LAS software. UV, FITC, Texas Red, YFP filters.

Victor Plate Reader

Location: Room 2032 Level 2 (WMI)

Contact: Hong Yu (59515; hong_yu@wmi.usyd.edu.au)

Description: A Wallac Victor 2 (1420) multi label counter. It has 7 filters for absorbance, and fluorescence filters.

Zeiss Elispot microscope (Elispot)

Location: Room 2032 Level 2 (WMI)

Contact: Hong Yu (59515; hong_yu@wmi.usyd.edu.au)

Description: Axioplan2 microscope for elispot counting. Equipped with HV-C20A HITACHI 3CCD camera, motorised stage, only one 5X objective. Fluorescence is not available. Counting software: KE elispot.

Kodak Imaging Station 4000MM

Location: Room 2032 Level 2 (WMI)

Contact: Hong Yu (59515; hong_yu@wmi.usyd.edu.au)

Description: Multi-wavelength illumination system allows fluorescence and bright field imaging. 10x optical zoom digital camera. Excitation filters: 465nm, 535nm; Emission filter: 535nm,600nm.

Microarray Scanner

Location: Gene expression lab Room 3004 Level 3 (WMI)

Contact: Ilya Henner (59000)

Description: The GenePix 4000B microarray scanner is a dual-wavelength laser scanner that produces high resolution images used in the acquisition of expression data from DNA microarrays, protein microarrays, tissue arrays and cell arrays.

Flow Cytometer FACSCALIBUR

Location: Flow room Level 2, Rm 2013 (WMI)

Contact: Mary Sartor (56257 or 56275; Mary@icpmr.wsahs.nsw.gov.au), or Sanda Lum (59050; sanda_lum@wmi.usyd.edu.au)

Description: 4 colour acquisition FACSCALIBUR Benchtop flow cytometer. 488nm and 633 nm lasers. Bookings required (2 hour limit per user). Bench top flow cytometer

Flow Cytometer FACSCAN

Location: ICPMR, Rm 2071

Contact: Mary Sartor (56257 or 56275; Mary@icpmr.wsahs.nsw.gov.au), or Sanda Lum (59050; sanda_lum@wmi.usyd.edu.au)

Description: 3 colour acquisition FACSCAN. 488nm laser. Bench top flow cytometer. Bookings are required

FACSDIVA cell sorter

Location: WMI flow room 2013

Contact: Mary Sartor (56257 or 56275; Mary@icpmr.wsahs.nsw.gov.au), or Sanda Lum (59050; sanda_lum@wmi.usyd.edu.au)

Description: 488nm and 635nm HeNe laser. The sorter is now FACSDIVA capable of high speed cell sorting and can acquire six colours. The facility is also PC2 compliant.

CELLQuest Workstation Locations:

WMI: Flow Room 2013 and Level 3- Rooms 3014 and 3013.

ICPMR: Flow Room 2071 and ICPMR: Renal dept

Children's Hospital: Research Building-Level 3 Microscopy Room, Rm 434876

Children's Medical Research Institute (CMRI)**Confocal (1990-now rarely used)**

Location: CMRI

Contact: Christine Smyth (9687 2800)

Description: Leica upright (circa 1990, upgraded 1996). Ar Krypton laser; excitation lines 488nm and 568nm for FITC, Liss Rh, Texas Red etc. CLSM software) S-9 v2.4.5.

FacsCanto Becton Dickinson

Location: CMRI

Contact: Christine Smyth (9687 2800)

Description: Ar ion laser and 633nm diode laser; Diva Software.

Beta-Plate Counter

Location: Room F26 (CMRI)

Contact: Greg Craig (GCraig@cmri.usyd.edu.au), Doreen Molasky, Grant Logan.

Description: For counting Beta Radiation in a 96 well plate format.

Storm System

Location: Room F27A (CMRI)

Contact: Greg Craig (GCraig@cmri.usyd.edu.au), Doreen Molasky.

Description: For quantitation of autoradiographs utilizing phosphor imaging screens and storm scanner.

Fluorescent Spectrophometer

Location: Room G 26 (CMRI)

Contact: Greg Craig (GCraig@cmri.usyd.edu.au), Doreen Molasky. Description: For the analysis/quantitation of fluorescently active samples.

UV/Vis Spectrophometer

Location: Room F27A (CMRI)

Contact: Greg Craig (GCraig@cmri.usyd.edu.au), Doreen Molasky (Lab Manager)

Description: For the analysis/quantitation of UV/Vis active samples.

Inverted Fluorescent Microscope

Location: Room G25 (CMRI)

Contact: Christine Smyth (csmyth@cmri.usyd.edu.au), Axel Neumann (aneumann@cmri.usyd.edu.au)

Description: Leica fluorescent microscope with Blue, Green and DAPI excitation filters. Spot b/w digital camera.

Upright Fluorescent Microscope

Location: Room F75 (CMRI)

Contact: Christine Smyth (csmyth@cmri.usyd.edu.au), Axel Neumann (aneumann@cmri.usyd.edu.au)

Description: Leica fluorescent microscope with Blue, Green and DAPI excitation filters. Spot colour digital camera.

Upright Fluorescent Microscope

Location: Room F75 (CMRI)

Contact: Christine Smyth (csmyth@cmri.usyd.edu.au), Axel Neumann (aneumann@cmri.usyd.edu.au)

Description: Olympus BX51 fluorescent microscope with Blue, Green and DAPI excitation filters. Spot b/w digital camera.

Motorised Inverted Fluorescent Microscope for in vivo imaging.

Location: Room G27 (CMRI)

Contact: Andrew McGeachie (amcgeachie@cmri.com.au)

Description: Olympus IX81 motorised inverted fluorescent microscope with motorised stage and image collection and analysis software. Hamamatsu Photonics Cooled CCD camera. Blue and Green filter sets.

InCell Scanner.

Location: Room G27 (CMRI)

Contact: Andrew McGeachie (amcgeachie@cmri.com.au)